Bioactivity-guided Isolation and Identification of Xanthine Oxidase Inhibitors from Morus alba Bark

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ABSTRACT

Objectives: The objective of this research was isolation and evaluation of xanthine oxidase inhibitory effects of isolated substances from mulberry bark (Morus alba L.). Methods: All four fractions: n-hexane, chloroform, ethyl acetate and water were evaluated for their effects on xanthine oxidase activity. The active compounds were isolated from the most potential fraction using column chromatography. The structures of these compounds were elucidated using ¹H and ¹³C-NMR and their xanthine oxidase inhibitory activities were evaluated spectrophotometrically. The pharmacokinetics parameters of the most promising substances were predicted using SwissADME tool. Results: Three compounds (Me01, Me02, Me04): ursolic acid Me01, oxyresveratrol Me02 and kuwanon G Me04 were isolated from ethyl acetate fraction. The xanthine oxidase activity assay showed that two compounds Me02 (oxyresveratrol) and Me04 (kuwanon G) both demonstrated inhibitory effects on xanthine oxidase with the IC₅₀ values of 42.08 µg/ml and 52.41 µg/ml, respectively. The predicted adsorption, distribution, metabolism and excretion (ADME) properties illustrated that oxyresveratrol (Me02) had high solubility in water, high gastro-intestinal (GI) absorption, no violation of Lipinski’s rule of five and was not affected by P-glycoprotein, which is the cause of poor bioavailability of many drugs. Conclusion: Our findings suggest that oxyresveratrol is potent natural xanthine oxidase inhibitor in drug discovery and development for prevention and treatment of gout.

Keywords: In vitro; Isolation; Morus alba; Mulberry bark; Xanthine oxidase

INTRODUCTION

Gout is a disorder of purine metabolism that increases the level of uric acid in the body, which is characterized by severe pain and swelling. This often has serious consequences for patients such as knee damage, joint deformities, kidney failure, if not diagnosed promptly and treated properly¹. Xanthine oxidase (XO) is an enzyme that plays an important role in the degradation of purine into uric acid in humans. XO enzyme inhibitors reduce uric acid levels in the body and are one of the drugs used to prevent and treat diseases related to hyperuricemia, including gout¹. However, the XO inhibitors such as allopurinol and febuxostat still have some undesirable effects including skin rash, nausea, vomiting, kidney failure, Steven - Johnson syndrome¹. Therefore, searching for herbs and natural compounds that have

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effect on xanthine oxidase activity with low toxicity has been recently considered as an alternative solution in the discovery and development of new drug for gout treatment.

In Vietnamese traditional medicine, mulberry (Morus alba L.) leaves (tang diep), fruits (tang tham), twigs (tang chi) and root bark (tang bach bi) are all valuable medicines which are used to treat many diseases such as headache, dizziness, hypertension, cough, osteoarthritis pain...2. Several researches have shown that mulberry twig extract reduces serum uric acid and increases uric acid excretion in white mice with hyperuricemia3,4. Although the trunk is one of the most similar parts to the twig in terms of its chemical composition, the relationship between the chemical composition and the in vitro inhibitory effect of in vitro xanthine oxidase of Morus alba L. trunk extract has not been known yet. Therefore, the study was performed with the aim of discovering potential xanthine oxidase inhibitors from mulberry tree trunks.

**MATERIAL AND METHODS**

**Plant materials**

The mulberry trunks (Morus alba L.) were collected in Song Mai, Bac Giang, Vietnam in August 2016. A plant sample was identified by Dr. Nghiem Duc Trong. The voucher specimen (ID: HNIP/18484/17) was deposited at the Herbarium of Department of Botany, Hanoi University of Pharmacy. The bark and the wood were separated, dried, cut into 1-2 cm pieces and placed in plastic bags.

**Chemicals**

Sephadex LH-20 (Sigma-Aldrich), normal phase silica gel 60 (0.040-0.063 mm; 230 - 400 mesh; Merck), reversed-phase silica gel (30-50 µm; Fujisilisa Chemical Ltd., Japan), thin layer chromatography DC-Alufolien 60 F254 (Merck), reversed-phase RP-18 F254 (Merck), vanillin-sulphuric acid spraying reagent for separation; DMSO (Sigma Aldrich), xanthine oxidase (Sigma Aldrich), xanthine (Sigma Aldrich), quercetin (National Institute of Drug Quality Control) for xanthine oxidase assay.

**Extraction and isolation**

The plant materials (500 g bark and 500 g wood) were separately ground and refluxed with methanol (the ratio of solid : solvent = 1: 6) thrice for 3 hours each. Then, the extracts were combined and evaporated to dryness under reduced pressure to afford 29.16 g and 38.84 g methanol extract from mulberry wood and bark, respectively. Next, the methanol extract was dispersed in hot water at the ratio of 1: 4, then liquid-liquid phase separated in turn with n-hexane, chloroform and ethyl acetate. Each fraction was concentrated under reduced pressure and dried in order to obtain n-hexane (11.20 g and 1.84 g), chloroform (3.27 g and 0.63 g), ethyl acetate (4.24 g and 2.74 g) and water fractions (7.63 g and 8.32 g) from mulberry bark and wood, respectively.

The fraction that exhibited the highest in vitro xanthine oxidase inhibitory activity was selected for fractionation and isolation of active constituents using silica gel column chromatography (particle size: 0.04 – 0.063 mm) with gradient eluting solvent dichloromethane (DCM) and methanol (MeOH) (1:0, 1:1, 0:1, v/v), yielding 12 fractions (F1 to F12). Me01 was isolated from fraction F1 by natural evaporation of solvent and recrystallization with ethanol. Me02 was isolated from fraction F7 by Sephadex LH-20 chromatography and eluted by methanol – water (5:1, v/v) as pale-yellow crystal while Me04 was obtained from fraction F10 using reversed-phase silica gel chromatography (particle size: 30 – 50 µm) with methanol – water (1:1, v/v) as mobile phase.

The structures of isolated compounds were elucidated based on the 1H and 13C-NMR spectral data which were recorded on a Bruker Avance 500 MHz for 1H and 125 MHz for 13C spectrometer.

**Xanthine oxidase assay**

The xanthine oxidase inhibitory activity of fractions and isolated compounds were assayed spectrophotometrically using method described by Mai et al. with modifications5. The reaction mixture consisting of 50 µl of test solution at different concentrations prepared by diluting 10 mg/ml solution of crude extract in dimethyl sulfoxide (DMSO) with 70 mM phosphate buffer (pH = 7.5), 35 µl of phosphate buffer and 30 µl of enzyme solution in phosphate buffer were incubated at 25°C for 15 minutes followed by addition of 60 µl of 150 µM xanthine and incubating for 30 minutes at 25°C. 30 µl of 1N HCl was added to stop the reaction and the absorbance was measured at 290 nm. Each experiment was performed in triplicate. Quercetin was used as positive control. The half maximal inhibitory concentration (IC50) values were calculated by using GraphPad Prism 8.0 software.

**Molecular docking and ADME prediction**

The crystal structures of xanthine oxidase (ID: 3NVY) were collected from RCSB protein data bank6. After removing water molecules, hydrogen atoms were added followed by protonating using Protonate3D process in MOE 2009 software. The partial charges were computed by Amber 99 forcefield. The structures of ligands were drawn using Chem Office 17.0 software then optimized and assigned atomic charges. After that, enzyme and ligands were docked, using Triangle Matcher method and London dG scoring function.
Table 1. Evaluation of *in vitro* xanthine oxidase inhibition effect of extracts and isolated compounds from mulberry trunk

<table>
<thead>
<tr>
<th>Part</th>
<th>Fraction/Compound</th>
<th>IC₅₀ (μg/ml)</th>
<th>95% Confidence Interval (μg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bark</td>
<td>Methanol</td>
<td>84.94</td>
<td>38.17 – 189.00</td>
</tr>
<tr>
<td></td>
<td>n-Hexane</td>
<td>145.10</td>
<td>58.51 – 359.70</td>
</tr>
<tr>
<td></td>
<td>Chloroform</td>
<td>151.50</td>
<td>92.80 – 247.20</td>
</tr>
<tr>
<td></td>
<td>Ethyl acetate</td>
<td>18.59</td>
<td>10.70 – 26.28</td>
</tr>
<tr>
<td></td>
<td>Water</td>
<td>&gt; 300</td>
<td>-</td>
</tr>
<tr>
<td></td>
<td>Methanol</td>
<td>125.40</td>
<td>38.78 – 405.60</td>
</tr>
<tr>
<td></td>
<td>n-Hexane</td>
<td>&gt; 300</td>
<td>-</td>
</tr>
<tr>
<td>Wood</td>
<td>Chloroform</td>
<td>47.19</td>
<td>41.23 – 56.04</td>
</tr>
<tr>
<td></td>
<td>Ethyl acetate</td>
<td>33.19</td>
<td>21.06 – 52.31</td>
</tr>
<tr>
<td></td>
<td>Water</td>
<td>&gt; 300</td>
<td>-</td>
</tr>
<tr>
<td>Positive control</td>
<td>Quercetin</td>
<td>2.77</td>
<td>1.81 – 4.03</td>
</tr>
<tr>
<td>Isolated compound</td>
<td>Ursolic acid (Me01)</td>
<td>&gt; 300</td>
<td>-</td>
</tr>
<tr>
<td>Isolated compound</td>
<td>Oxyresveratrol (Me02)</td>
<td>42.08</td>
<td>55.34 – 76.57</td>
</tr>
<tr>
<td>Isolated compound</td>
<td>Kuwanon G (Me04)</td>
<td>52.41</td>
<td>38.03 – 72.75</td>
</tr>
</tbody>
</table>

![Chemical structure](http://aprh.journals.ekb.eg/)  

**Figure 1.** The structure of isolated compounds. **A.** Ursolic acid; **B.** Oxyresveratrol; **C.** Kuwanon G
ADME parameters of isolated compounds were predicted using Swiss ADME, a free web tool. The predicted properties included water solubility, GI absorption, blood-brain barrier (BBB) permeant, interaction with P-glycoprotein (P-gp), cytochromes P450 (CYPs) inhibition and number of violations of Lipinski’s rule of five of ligands.

**RESULTS**

**Isolation and structure elucidation**

The effect of methanol extracts and four fractions (n-hexane, chloroform, ethyl acetate and water) of mulberry bark and wood on xanthine oxidase activity were evaluated at six concentrations: 3, 10, 30, 50, 100 and 300 µg/ml. The results are presented in Table 1.

As shown in Table 1, the extract from mulberry bark had a better inhibitory effect on xanthine oxidase activity than the core extract. On the other hand, the ethyl acetate fraction showed the best inhibitory activity with IC50 of 18.59 µg/ml. Therefore, the ethyl acetate fraction from mulberry bark was chosen to isolate potential xanthine oxidase inhibitors.

Using silica gel column chromatography, Sephadex LH-20 chromatography and reversed-phase silica gel chromatography, three pure compounds, Me01, Me02 and Me04 were isolated. Their structures were elucidated based on the NMR spectral data. Me01 was obtained as white amorphous powder. 1H-NMR and 13C-NMR spectra data of Me01 were represented below:

- **1H-NMR** (500 MHz, CDCl3 & CD2OD), δ (ppm): 5.24 (1H, t, J = 7.5 Hz, H-12), 3.20 (1H, t, J = 16.5 Hz, H-3), 2.19 (1H, d, J = 11.0 Hz, H-18), 1.09 (3H, s, H-27), 0.98 (3H, s, H-26), 0.95 (3H, s, H-25), 0.93 (3H, s, H-30), 0.86 (3H, d, J = 6.5 Hz, H-29), 0.81 (3H, s, H-23), 0.77 (3H, s, H-24).

- **13C-NMR** (125 MHz, CDCl3 & CD2OD), δ (ppm): 180.7 (C-28), 138.0 (C-13), 125.4 (C-12), 78.8 (C-3), 55.1 (C-5), 52.7 (C-18), 47.7 (C-17), 47.4 (C-9), 41.9 (C-14), 39.4 (C-8), 38.9 (C-19), 38.8 (C-20), 38.6 (C-1), 38.5 (C-4), 36.8 (C-22), 36.7 (C-10), 32.9 (C-7), 30.6 (C-21), 29.6 (C-15), 27.9 (C-23), 26.7 (C-2), 24.1 (C-16), 23.4 (C-27), 23.2 (C-11), 21.0 (C-30), 18.2 (C-6), 16.9 (C-26), 16.8 (C-29), 15.5 (C-24), 15.3 (C-25).

The 1H-NMR and 13C-NMR spectra showed the presence of the proton of the hydroxymethine group at 3.20 ppm (1H, t, J = 16.5 Hz, H-3) corresponding to the signal of carbon C-3 at the δc of 78.8 ppm. The presence of olefinic hydrogen was confirmed by signals at 5.24 ppm (1H, t, J = 7.5 Hz, H-12). Based on the 13C-NMR and DEPT spectra, it could be concluded that there were 30 carbons, including 7 primary carbons, 9 secondary carbons, 7 tertiary carbons and 7 quaternary carbons. By comparison with published spectral data, Me01 was identified as ursolic acid (Figure 1A).

Compounds Me02 was obtained as needle-shaped and light greenish yellow crystal. 1H-NMR and 13C-NMR spectra data of Me02 were represented below:

- **1H-NMR** (500 MHz, Acetone-d6), δ (ppm): 6.23 (1H, t, J = 2 Hz, H-4), 6.37 (1H, dd, J = 8.5 Hz, 2.5 Hz, H-5), 6.43 (1H, d, J = 2.5 Hz, H-3'), 6.51 (2H, d, J = 2 Hz, H-2, H-6), 6.88 (1H, J = 16.5 Hz, H-8), 7.32 (1H, d, J = 16.5 Hz, H-7), 7.39 (1H, d, J = 8.5 Hz, H-6').

- **13C-NMR** (125 MHz, Acetone-d6), δ (ppm): 159.6 (C-3, C-5), 159.1 (C-4'), 156.9 (C-2'), 141.6 (C-1), 128.2 (C-6'), 126.3 (C-8), 124.4 (C-7), 117.3 (C-1'), 108.4 (C-5'), 105.5 (C-2, C-6), 103.6 (C-3'), 102.3 (C-4)

13C-NMR spectrum exhibited 14 signals corresponding to 14 carbons, including four aromatic ring carbons linked to hydroxy appearing at δc 159, 159.6 ppm (C-3', C-5'), 159.1 ppm (C-4') and 156.9 ppm (C-2'), 10 sp3 hybridized carbons signals in the range of δc 102.3 to 141.6 ppm; The DEPT spectrum showed that there are 6 quaternary carbons and 8 primary carbons. Based on the 1H-NMR, 13C-NMR spectra and published data, the structure of Me02 were identified as oxyresveratrol (Figure 1B).

Compounds Me04 is reddish-brown, amorphous powder. 1H-NMR and 13C-NMR spectra data of Me04 were represented below:

- **1H-NMR** (500MHz – Acetone-d6): 7.42 (1H, H-27), 7.25 (1H, H-6'), 6.77 (1H, d, J = 8.5 Hz, H-33), 6.69 (1H, brs, H-3'), 6.56 (1H, d, J = 8.5 Hz, H-7'), 6.21 (1H, brs, H-30), 6.07 (1H, d, J = 8.5Hz, H-32), 6.04 (1H, s, H-6), 5.98 (1H, brs, H-24), 5.96 (1H, d, J = 8.5 Hz, H-26), 5.18 (2H, m, H-10, H-15), 4.44 (1H, d, H-19), 3.32 -4.43 (2H, m, H-14, H-20), 3.21 (2H, H-9), 1.70 -1.98 (2H, m, H-18), 1.61 (3H, s, H-12), 1.52 (3H, s, H-17), 1.48 (3H, s, H13).

- **13C-NMR** (125MHz – Acetone-d6): 206.6 (C-21), 183.1 (C-4'), 165.6 (C-23), 164.9 (C-25), 162.1 (C-7), 161.5 (C-4'), 161.3 (C-8a), 160.8 (C-2), 157.3 (C-2'), 157.1 (C-5), 157.1 (C-29), 157.1 (C-31), 133.9 (C-27), 133.6 (C-16), 132.1 (C-6'), 132.1 (C-33), 132.1 (C-11), 124.4 (C-15), 122.9 (C-10), 121.4 (C-28), 115.6 (C-3), 113, 2 (C-1'), 107.9 (C-5'), 107.9 (C-32), 107.9 (C-26), 107.6 (C-22), 105.4 (C-8 ), 103.6 (C-3'), 103.6 (C-30), 103.5 (C-4a), 102.8 (C-24), 98.3 (C-6), 47.6 (C-20), 38.6 (C-19), 25.8 (C-12), 24.5 (C-14), 24.5 (C-18), 24.5 (C-9), 23.1 (C-17), 17.7 (C-13).

The 1H-NMR (500MHz – Acetone-d6) spectrum of Me04 showed the presence of three ABX spin systems with δH (ppm): 7.25 (1H, H-6'), 6.56 (1H, d, J = 8.5Hz, H-5') and 6.69 (1H, brs, H-3'), 5.96 (1H, d, J = 8.5 Hz, H-26), 7.42 (1H, H-27) and 5.98 (1H, brs, H-24); 6.77 (1H, d, J = 8.5, H-33), 6.07
Table 2. Predicted ADME properties of oxyresveratrol (Me02) and kuwanon G (Me04)

<table>
<thead>
<tr>
<th>Compound</th>
<th>Water solubility</th>
<th>GI absorption</th>
<th>BBB permeant</th>
<th>P-gp substrate</th>
<th>CYP inhibition</th>
<th>Lipinski’s rule of five</th>
</tr>
</thead>
<tbody>
<tr>
<td>Me02</td>
<td>Soluble</td>
<td>High</td>
<td>No</td>
<td>No</td>
<td>CYP1A2, CYP2C9, CYP3A4</td>
<td>0 violation</td>
</tr>
<tr>
<td>Me04</td>
<td>Poorly soluble</td>
<td>Low</td>
<td>No</td>
<td>No</td>
<td>No</td>
<td>3 violations</td>
</tr>
</tbody>
</table>

GI: Gastro-Intestinal, BBB: Blood-Brain Barrier, P-gp: P-glycoprotein, CYP: cytochrome P450

Figure 2. The binding mode of kuwanon G (A) and oxyresveratrol (B) with xanthine oxidase

Evaluation of xanthine oxidase inhibitory activity of isolated compounds

The effect of ursolic acid, oxyresveratrol and kuwanon G on xanthine oxidase activity in vitro was evaluated and presented in Table 1. The results showed that only oxyresveratrol and kuwanon G exhibited xanthine oxidase inhibitory effect with IC₅₀ values of 42.08 µg/ml and 52.41 µg/ml, respectively. In order to investigate the mechanism of action of oxyresveratrol and kuwanon G, molecular docking was performed. The results were shown in Figure 2.

As shown in Figure 2, the hydrogen bonding between hydroxyl group of oxyresveratrol, kuwanon G and Glu 802, Arg880, Thr1010 residues and π-π interaction between phenyl moiety and Phe914 residue of xanthine oxidase might play key role in XO inhibitory activity. These results were consistent with researches of C. M. Lin¹¹ and S. Lin¹², in which, they identified that flavonoids occupied the active cavity of XO by interaction with Glu802, Arg880, Phe914 and Thr1010, four of the key residues in the active site of enzyme.

Prediction of ADME parameters is an important step in optimization of lead compound. In this study, water solubility, GI absorption, BBB permeant, interaction with P-gp, CYP inhibition and number of violations of Lipinski’s rule of five of oxyresveratrol (Me02) and kuwanon G (Me04) were estimated using SwissADME tool and showed in Table 2.

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The results in Table 2 illustrated that oxyresveratrol (Me02) was a potential candidate in drug research and development of xanthine oxidase inhibitor with high solubility in water, high GI absorption and no violation of Lipinski’s rule of five. The results also indicated that oxyresveratrol was not affected by P-glycoprotein, which is the cause of poor bioavailability of many drugs.

**Discussion**

According to the results of evaluating the *in vitro* xanthine oxidase inhibitory effect of methanol extracts and fractions, the ethyl acetate fraction from mulberry trunk bark showed the strongest activity with IC$_{50}$ of 18.59 µg/ml. Ethyl acetate is a slightly polar solvent with Polarity Index of 4.4, therefore this solvent could well dissolve most of flavonoids. It can be predicted that this class of compounds might play a key role in XO inhibitory effect of ethyl acetate extract from mulberry barks. From the ethyl acetate fraction, ursoic acid, a pentacyclic triterpenoid carboxylic acid and two other compounds with similar framework, oxyresveratrol and kuwanon G, were isolated. However, only oxyresveratrol and kuwanon G demonstrated effect on xanthine oxidase activity with IC$_{50}$ values of 42.08 µg/ml and 52.41 µg/ml, respectively.

Oxyresveratrol is a stilbene derivative presented in many species such as *Smilax china* L. (*Smilacaceae*)$^{13}$, *Morus alba* L. (*Moraceae*)$^{14}$, *Morus nigra* L. (*Moraceae*)$^{15}$. Previous studies have demonstrated that oxyresveratrol could inhibit the inflammatory response *in vitro* and *in vivo* - one of the typical symptoms of acute gout$^{16}$, prevent DNA damage more effective than both trolox and ascorbic acid$^{17}$, inhibit protein kinase C$^{18}$ or reduce the production of TNF-α necrosis factor$^{19}$. In this study, oxyresveratrol was considered as a potential compound in prevention and treatment of gout with high affinity toward xanthine oxidase and suitable ADME properties. However, more studies are still required in order to confirm the effectiveness of oxyresveratrol clinically.

Kuwanon G is a common flavonoid Diels-Alder that could be easily found in *Morus* species$^{20, 21}$. Recent studies have shown that kuwanon G could relieve the symptoms of asthma$^{22}$, decrease the disruption of the intestinal epithelium by improving the cell viability and tight junction activity, reduce the production of inflammatory cytokines and prevent oxidative damage$^{23}$.

This is the first study in which the *in vitro* xanthine oxidase inhibitory activity of oxyresveratrol and kuwanon G was assessed. The molecular docking results revealed that these two compounds competitively inhibited XO activity by forming the hydrogen bonding with Glu 802, Arg880, Thr1010 residues of xanthine oxidase, which were consistent with researches reported by C. M. Lin$^{11}$ and S. Lin$^{12}$. These are remarkable features in drug discovery and development for prevention and treatment of gout.

**Conclusion**

From mulberry trunk bark (*Morus alba* L.), three compounds Me01, Me02 and Me04 were isolated. Based on the nuclear magnetic resonance spectroscopy data, it was identified that Me01 was ursoic acid, Me02 was oxyresveratrol and Me04 was kuwanon G. The xanthine oxidase assay showed that two compounds Me02 (oxyresveratrol) and Me04 (kuwanon G) have inhibitory effects on xanthine oxidase activity with IC$_{50}$ values of 42.08 µg/ml and 52.41 µg/ml, respectively. The predicted ADME properties suggested that oxyresveratrol was a potent lead in novel drug discovery for gout. However, more studies are needed in order to confirm the effectiveness of oxyresveratrol clinically.

**Conflict of interest**

The authors declare that there is no conflict of interest regarding the publication of this paper.

**References**


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